

1 **Global dissemination patterns of common gene cassette arrays in class 1 integrons**

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Abbreviations: MGEs, mobile genetic elements; 5'-CS, 5'-conserved segment; 3'-CS, 3'-conserved segment; ISCR1, insertion sequence common region; HGT, horizontal gene transfer; orf, open reading frame.

27 **Summary**

28 Integrons are genetic elements that contain a site-specific recombination system able to
29 capture, express and exchange gene cassettes. Mobile integrons are widespread and
30 often confer resistance to multiple antibiotics, due to the expression of the arrays of
31 gene cassettes they carry. Although more than 300 cassette arrays have been described,
32 less than 10 array compositions prevail in the reports related with class 1 integrons.
33 These common arrays are found in a broad variety of hosts and environments,
34 highlighting the high level of horizontal dissemination of these elements among
35 bacterial populations and species. Clonal expansion also contributes to the current
36 prevalence of and inter-regional spread of integron-carrying bacterial species. Here we
37 review the dissemination pattern of common cassette arrays with a focus on the
38 bacterial species, the geographical dispersal pattern and the environments in which they
39 reside. Conserved arrays of gene cassettes are found in at least 74 countries and 72
40 species present in different environments. The factors governing the further spread and
41 population dynamics of these cassette arrays remain to be determined.

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61 **Introduction**

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63 Integrins are genetic elements that contain a site-specific recombination system able to
64 capture, express and exchange specific DNA elements, called gene cassettes (Hall &
65 Collis, 1995). Gene cassettes are genetic units that include a single *open reading frame*
66 (orf), usually without promoter, and a specific recombination site, *attC* (Domingues *et*
67 *al.*, 2012a; Recchia & Hall, 1995).

68 Integrins are present in approx. 10% of the bacterial genomes sequenced, and
69 contribute to genomic plasticity and environmental adaptation of bacteria (Boucher *et*
70 *al.*, 2007). In fact, integrins can be considered one of the major genetic carriers of and
71 vectors for dissemination of antibiotic resistance determinants in bacteria (Cambray *et*
72 *al.*, 2010). Integrins have been classified according to their genomic context as either
73 mobile integrins, whenever they are associated with transposons, or chromosomal
74 integrins, when located in the bacterial chromosome (Cambray *et al.*, 2010; Mazel,
75 2006). Mobile integrins are mainly found in Gram-negative bacteria, though a few
76 studies have also reported their presence among Gram-positive species (Cambray *et al.*,
77 2010; Domingues *et al.*, 2012a). The genetic relatedness of the integrase gene of the
78 mobile integrins is used to group them into distinct classes (Boucher *et al.*, 2007;
79 Stokes & Hall, 1989). Based on the nucleotide sequence of the integrase, five classes of
80 mobile integrins have been described (Cambray *et al.*, 2010). Class 1 integrins were
81 described in 1989 (Stokes & Hall, 1989), and are reported as the most common and
82 widespread, especially in clinical settings. This class mainly carries antimicrobial
83 resistance genes and it is highly disseminated due to the close association with
84 transposons, often embedded in conjugative plasmids (Cambray *et al.*, 2010).

85 All integrins consist of three essential elements, the *intI* integrase gene, which encodes
86 the IntI protein responsible for the site-specific recombination of gene cassettes, the
87 adjacent recombination site *attI* and the gene cassette promoter P_c (previously called
88 P_{ANT}) (Hall & Collis, 1998), although the presence of the promoter has not been shown
89 in all classes (Boucher *et al.*, 2007). An additional promoter, P_2 , is present in some
90 integrins (Collis & Hall, 1995).

91 The complete integron is not considered to be a mobile element itself as it lacks
92 functions for self-mobility. However, the individual gene cassettes are considered
93 mobile within integrins (Mazel, 2006). Furthermore, integrins are often embedded in
94 larger mobile genetic elements (MGEs) such as transposons and plasmids which

95 mediate horizontal dissemination of the integrons (Domingues *et al.*, 2012a; Fluit &
96 Schmitz, 1999; Gombac *et al.*, 2002). The functions, abundance and selection of MGEs
97 in bacterial communities are therefore important for further integron dispersal and
98 bacterial evolution (Aminov, 2011; Wright *et al.*, 2008). This ability of broad and rapid
99 dispersal has critical clinical implications, as the majority of the gene cassettes in
100 examined mobile integrons encode antimicrobial resistance (Partridge *et al.*, 2009;
101 Recchia & Hall, 1995).

102 Integrons with identical gene cassette arrays are found in different species from diverse
103 environments and geographic locations. Here, we review the occurrence and
104 dissemination pattern of some of the most common gene cassette arrays (Partridge *et*
105 *al.*, 2009) found in mobile integrons belonging to class 1.

106

107 **Class 1 integrons**

108 Class 1 integrons have usually three distinct genetic regions: two highly conserved
109 regions, the 5'-conserved segment (5'-CS) and the 3'-conserved segment (3'-CS),
110 flanking the central but variable region where the gene cassettes are located (Stokes &
111 Hall, 1989); these integrons have been designated classic class 1 integrons. The 5'-CS
112 includes the *intII* gene, the *attII* site and the promoters P_c, and P2 when present. The 3'-
113 CS consists of the *qacEΔ1* gene, which encodes an incomplete version of a protein that
114 mediates resistance to certain detergents, the *sul1* gene, encoding resistance to
115 sulphonamides, and an open reading frame, *orf5*, of unknown function (Figure 1). The
116 central variable region can have different numbers and types of gene cassettes resulting
117 in integrons with diverse compositions of gene cassette arrays. The variable region of
118 class 1 integrons is often targeted in PCR with a pair of primers called 5'-CS and 3'-CS,
119 which bind in the conserved regions of this class (Levesque *et al.*, 1995). The size of the
120 amplicon of each cassette array described in this manuscript refers to the PCR product
121 obtained with this primer pair. It is noted that several other primer pairs have been
122 developed and the PCR amplicon produced by these can have different sizes although
123 they target the same arrays (Ajiboye *et al.*, 2009; Sandvang *et al.*, 1998; White *et al.*,
124 2000).

125 Although this is the most commonly described structure of class 1 integrons, other
126 structures have been identified: such as the integrons without a 3'-CS (Dawes *et al.*,
127 2010; Pan *et al.*, 2006), and others with more complex structures. The term complex
128 class 1 integrons has been used for the ones that includes ISCR1 (Insertion Sequence

129 Common Region), partial duplications of the 3'-CS and an addition ISCR1-associated
130 region that usually carries a non-cassette resistance gene (Bennett, 2008; Toleman *et al.*,
131 2006a, b).

132

133 **Gene cassettes**

134 Numerous combinations of gene cassettes have been reported (Partridge *et al.*, 2009). A
135 number is assigned to each new integron based on the cassette array and its unique
136 nucleotide and amino acid sequence. A list of numbered class 1 integrons is available
137 from the INTEGRALL database (<http://integrall.bio.ua.pt/>) (Moura *et al.*, 2009). So far
138 more than 1050 gene cassette arrays with single or multiple nucleotide differences have
139 been identified in class 1 integrons (database last accessed on 17 November 2014). It is
140 noted that the INTEGRALL database primary focuses on gene cassette compositions
141 whereas early integron numbers also indicate the structure beyond the last cassette
142 (Partridge *et al.*, 2002). Thus the taxonomy of integrons has changed over time and may
143 or may not consider both the gene cassettes and the characteristics of the flanking DNA
144 (e.g. defective Tn402 sequences). Unfortunately, the non-uniform taxonomy has also
145 led to a lack of full oversight in the designation of gene cassettes. For instance, some
146 identical gene cassette compositions appear in different studies with different
147 designations, as the same gene has been named differently. Additionally, the same name
148 has also been attributed to different genes. This lack of consistency can be confusing in
149 the case of naming gene cassettes of unknown function. A new nomenclature for these
150 *orf* cassettes was proposed. The designation *gcu* (gene cassette of unknown function)
151 was proposed to refer to cassettes with an *attC* site but with unknown function
152 (Partridge *et al.*, 2009). Some examples of inconsistencies in the naming of gene
153 cassettes are given in Table 1 (Du *et al.*, 2005; Partridge & Hall, 2005; Partridge *et al.*,
154 2009; Ramirez & Tolmasky, 2010). Furthermore, most of the published studies do not
155 provide the specific nucleotide sequence of the described genes. This practice also
156 hampers the correct identification of the arrays considered.

157

158 Common gene cassette arrays

159 According to Partridge and colleagues (Partridge *et al.*, 2009), the single gene cassettes
160 *aadB*, *dfrA7*, *aadA1a*, *aadA2*, *blaP1* [now called *bla*_{CARB-2} (www.lahey.org/Studies -
161 accessed on 17 November 2014)], and the arrays *dfrA1-gcuC*, *dfrA1-aadA1a*, *dfrA17-*

162 *aadA5* and *dfrA12-gcuF-aadA2* were the most common ones reported in class 1
163 integrons surrounded by the 5'-CS and 3'-CS regions (see further description below).

164 There is no standard system available to indicate nucleotide variants of gene cassettes.
165 For the purpose of this review, we assume, like these authors, that the gene cassettes are
166 only different (for the purpose of classification) if their nucleotide sequence differs by
167 more than 2%. An exception was made for *blaCARB-2* cassettes for which a single
168 nucleotide change, resulting in an amino acid change, lead to classification as a different
169 gene (www.lahey.org/Studies - accessed on 07 April 2015). Below we list key
170 characteristics of commonly occurring gene cassette arrays identified by Partridge and
171 colleagues (Partridge *et al.*, 2009), before a closer examination of their species,
172 geographical and environmental distribution.

173 Information on the distribution of each cassette array was collected from Pubmed,
174 GenBank and INTEGRALL databases using the designation of each gene cassette or
175 cassette array as the search term. In Pubmed, the term “integron” was added to the
176 search, together with each cassette/array. For each gene, additional search with the
177 alternative names given to the same cassette were performed. For example, for the
178 cassette array “*dfrA17-aadA5*” the searches performed in Pubmed were: “*dfrA17 AND*
179 *integron*”, “*dfr17 AND integron*”, “*dhfrXVII AND integron*”, “*aada5 AND integron*”,
180 “*ant(3')*-*1e AND integron*”, “*dfrA17-aadA5 AND integron*”, “*dfr17-aadA5 AND*
181 *integron*”, “*dhfrXVII-aada5 AND integron*”, “*dfrA17- ant(3')*-*1e AND integron*”,
182 “*dfr17- ant(3')*-*1e AND integron*”, and “*dhfrXVII- ant(3')*-*1e AND integron*”.

183 The references that are used as examples in this manuscript include studies that have
184 determined the cassette array by nucleotide sequence, size of the PCR amplicon and
185 restriction analysis, PCR amplification of the gene cassettes with specific primers,
186 and/or Southern blot hybridization, except for *blaCARB-2*, where only studies that
187 performed nucleotide sequence where included.

188

189

190 *aadB*: the *aadB* gene encodes an aminoglycoside 2"-adenylyltransferase conferring
191 resistance to dibekacin, gentamicin, kanamycin, sisomicin and tobramycin (Shaw *et al.*,
192 1993). The *aadB* gene cassette is 591 bp long (Recchia & Hall, 1995), and the PCR
193 amplicon is 744 bp. This gene cassette was first identified in the variable region of a
194 class 1 integron in an Australian clinical isolate of *Escherichia coli* (isolated in 1978)
195 and first described in 1986 (Cameron *et al.*, 1986; Stokes & Hall, 1989). This was later

196 shown to be embedded in the variable region of a complex class 1 integron, with
197 duplication of the 3'-CS (Hall & Stokes, 1990).

198

199 *dfrA7*: the *dfrA7* gene encodes a type VII dihydrofolate reductase, conferring
200 trimethoprim resistance (Roberts *et al.*, 2012; van Hoek *et al.*, 2011). The PCR
201 amplicon obtained with this cassette is 770 bp long, while the cassette itself is 617 bp
202 long (Recchia & Hall, 1995). The first report of this gene as part of a class 1 integron
203 dates to 1993 (Sundstrom *et al.*, 1993), when it was described to be located on plasmids
204 from *E. coli* isolated in Sweden in 1974 (Tennhammar-Ekman & Skold, 1979) and Sri
205 Lanka in 1981 (Sundstrom *et al.*, 1987).

206

207 *aadA1a*: the *aadA1a* gene encodes an aminoglycoside 3"-adenylyltransferase,
208 associated with resistance to spectinomycin and streptomycin (Ramirez & Tolmasky,
209 2010). The cassette is 856 bp long (Recchia & Hall, 1995). A class 1 integron with this
210 cassette will produce a PCR amplicon of 1009 bp. The *aadA1* gene has been associated
211 with the Tn21 transposon for many years (Liebert *et al.*, 1999); the first strain known to
212 carry it was *Shigella flexneri*, isolated in Japan in the late 1950s (Nakaya *et al.*, 1960).
213 In 1989, when integrons were first described, this gene was found to be associated with
214 a class 1 integron (Stokes & Hall, 1989).

215

216 *aadA2*: the *aadA2* gene encodes an aminoglycoside 3"-adenylyltransferase that confers
217 resistance to spectinomycin and streptomycin (Ramirez & Tolmasky, 2010). The *aadA2*
218 gene cassette is 856 bp long (Recchia & Hall, 1995) and the PCR amplicon is 1009 bp
219 long. The first report of a class 1 integron with this single cassette dates back to 1995,
220 and refers to a clinical isolate of *Pseudomonas aeruginosa* isolated in Japan in 1965
221 (Kazama *et al.*, 1995). Besides the classic class 1 integron, the widely disseminated
222 *aadA2* gene cassette is also present in complex class 1 integrons, such as in the
223 *Salmonella* resistance island SGI1 and some of the variants (Boyd *et al.*, 2002; Boyd *et*
224 *al.*, 2008).

225

226 *blaCARB-2*: the β-lactamase encoded by the *blaCARB-2* gene confers resistance to
227 penicillins including carbenicillin (Matthew & Sykes, 1977). The cassette is 1044 bp
228 (Recchia & Hall, 1995) and the PCR amplicon is 1197 bp long. The first report of the
229 integron-borne *blaCARB-2* gene cassette was published in 1991 in a plasmid from a *P.*

230 *aeruginosa* (Huovinen & Jacoby, 1991) isolate collected years before (Levesque &
231 Jacoby, 1988). The composition of this gene cassette is not always known in detail
232 (Soufi *et al.*, 2012; Van *et al.*, 2007). The *bla*CARB-2 cassette is often found adjacent to a
233 5'-CS fused to the *groEL* gene in the integron of the *Salmonella enterica* genomic
234 islands SGI1, which can also include ISCR1 (Boyd *et al.*, 2001; Doublet *et al.*, 2004). In
235 addition, this cassette can also be part of complex class 1 integrons, such as the one
236 inserted in the *S. enterica* islands SGI1-A and SGI1-G (Doublet *et al.*, 2004).

237

238 *dfrA1-gcuC*: the *dfrA1* cassette is a dihydrofolate reductase type I, conferring
239 trimethoprim resistance (Roberts *et al.*, 2012; van Hoek *et al.*, 2011); the *gcuC* cassette
240 is a hypothetical protein of unknown function. Whereas the gene cassette *dfrA1* is 577
241 bp long (Recchia & Hall, 1995), the *gcuC* is 512 bp long (Partridge *et al.*, 2009), and
242 the gene cassette array composed by these two cassettes produces a PCR amplicon of
243 1242 bp. This class 1 integron array was first reported in 1990 (amplicon size 1236 bp),
244 in the plasmid pLMO150 extracted from an Enterobacterium (Sundstrom & Skold,
245 1990). This particular cassette array can also be part of the genomic island SGI1, being
246 embedded in a complex class 1 integron in this case (Boyd *et al.*, 2008).

247

248 *dfrA1-aadA1a*: as previously mentioned, the *dfrA1* gene encodes a dihydrofolate
249 reductase type I, conferring resistance to trimethoprim (Roberts *et al.*, 2012; van Hoek
250 *et al.*, 2011); and the *aadA1a* gene encodes the enzyme aminoglycoside 3"-
251 adenylyltransferase associated with resistance to spectinomycin and streptomycin
252 (Ramirez & Tolmasky, 2010). The individual sizes of the cassettes *dfrA1* and *aadA1a*
253 are, as mentioned before, 577 bp and 856 bp, respectively (Recchia & Hall, 1995); the
254 *dfrA1-aadA1a* array PCR amplicon is 1586 bp long. The first report of this class 1
255 integron array dates from 1990, in the plasmid pLMO229 of an Enterobacterium
256 (Sundstrom & Skold, 1990)

257

258 *dfrA17-aadA5*: the *dfrA17* gene encodes a dihydrofolate reductase type XVII, also
259 conferring trimethoprim resistance (Roberts *et al.*, 2012; van Hoek *et al.*, 2011); an
260 aminoglycoside 3"-adenylyltransferase, associated with spectinomycin and
261 streptomycin resistance, is encoded by the *aadA5* gene (Ramirez & Tolmasky, 2010).
262 The PCR product obtained from the amplification of a classic class 1 integron
263 composed by the array *dfrA17-aadA5* is 1664 bp; the *dfrA17* and *aadA5* cassettes are

264 616 bp and 895 bp long, respectively (Partridge *et al.*, 2009). This gene cassette array
265 was reported for the first time in 2000, in the chromosome of a clinical *E. coli* isolate
266 collected in Australia in 1998 (White *et al.*, 2000). It is noted that the White et al. paper
267 does not suggests the *aadA5* gene confer streptomycin resistance. However, reports
268 studying the gene when present in other arrays suggest so (Sandvang, 1999).

269

270 *dfrA12-gcuF-aadA2*: the *dfrA12* gene encodes a type XII dihydrofolate reductase,
271 which confers trimethoprim resistance (Roberts *et al.*, 2012; van Hoek *et al.*, 2011); the
272 *gcuF* gene encodes a hypothetical protein of unknown function; and the *aadA2* gene, as
273 mentioned before, encodes an aminoglycoside 3"-adenylyltransferase, associated with
274 spectinomycin and streptomycin resistance (Ramirez & Tolmasky, 2010). The array
275 composed by these three gene cassettes produces a PCR amplicon of 1913 bp. The
276 *dfrA12* cassette is 584 bp long, the *gcuF* cassette is 320 bp long and the *aadA2* cassette
277 is 856 bp (Partridge *et al.*, 2009; Recchia & Hall, 1995). The first report of a classic
278 class 1 integron with this cassette array dates back to 1993, when it was identified in the
279 plasmid of a clinical *E. coli* isolate originally collected in Finland in the 1980's
280 (Heikkila *et al.*, 1993).

281

282 **Bacterial species distribution**

283 The nine commonly occurring gene cassette arrays (Partridge et al., 2009) considered in
284 this review have altogether been reported in at least 72 bacterial species (Table 2). They
285 are predominately found in class 1 integrons from Gram-negative bacterial species, as is
286 also the case for mobile integrons in general. However, a few arrays, specifically
287 *aadA1a*, *aadA2*, *dfrA17-aadA5*, and *dfrA12-gcuF-aadA2*, have also been detected in
288 Gram-positive bacteria.

289 Some bacterial species seem more likely to carry integrons than others. For instance,
290 more than six of the arrays have been found in bacteria belonging to the species
291 *Acinetobacter baumannii*, *E. coli*, *Klebsiella pneumoniae*, *Proteus mirabilis* and *S.*
292 *enterica*. In contrast, only one type of array has been described in the species
293 *Citrobacter braakii*, *Pseudomonas putida*, *Serratia liquefaciens* and *Vibrio*
294 *alginolyticus*. The species carrying the broader set of arrays are all known nosocomial
295 pathogens, often exposed to diverse antibiotic classes, and they are present in the same
296 environments. Thus, they are expected to have opportunities for physical contact and

297 horizontal exchange as well as being under strong positive selection for the resistance
298 traits encoded by the particular gene cassette compositions considered here.

299 Among the reviewed arrays, *aadA1a*, *aadA2* and *dfrA12-gcuF-aadA2* are present in a
300 diverse range of bacterial species (>30), while *aadB* and *blaCARB-2* are restricted to a
301 smaller number of species and seem to have preference for *Enterobacteriaceae* and *S.*
302 *enterica*, respectively.

303 Further studies should be conducted to better understand to what extent gene cassette
304 distribution patterns are limited by host species and environmental factors. As integrons
305 are frequently genetically linked to MGEs, consideration of the host ranges of linked
306 MGE and the impact of linked selection is essential in this context. See below for a
307 further discussion on limitations in studies due to biased or limited sampling or
308 publication effort.

309

310 **Geographical dissemination**

311 Class 1 integrons are globally disseminated. Class 1 integrons carrying the nine
312 common arrays considered here have been described in at least 73 countries (Table 3)
313 belonging to all continents, with the exception of Antarctica. However, some of the
314 gene cassette arrays considered appear (based on published literature) to have a
315 continent-related prevalence. For instance, the *dfrA1-gcuC* array is mostly reported in
316 Asia, while the *aadB* cassette is mainly reported in European countries. We can find
317 reports of all nine arrays in China, Malaysia, Tunisia and the United States. Other
318 countries, such as Austria and Zimbabwe, have so far reported bacteria with only one
319 type of the common arrays. If the geographical distribution pattern reflects a true
320 distribution or a biased or limited sampling or publication effort is yet not known.

321 More studies are needed to understand the causal factors determining the initial events
322 creating the particular gene cassette array as well as the population dynamic factors
323 resulting in the currently observed global dissemination patterns of antibiotic resistance
324 genes associated with class 1 integrons. Particularly, there is a need to better understand
325 the interplay between various population genetic processes (e.g. selection from
326 antibiotic usage, co-selection, vectors, genetic drift, dispersal) in determining further
327 geographic dissemination of these integrons. The effects of various antibiotic usage
328 levels and patterns in different countries must also be considered in this context.

329

330 **Environmental distribution**

331 As expected from the dissemination patterns of antimicrobial resistance in general,
332 resistance-encoding gene cassette arrays are mostly reported in bacteria isolated from
333 human clinical settings. Nonetheless, there are an increasing number of studies
334 describing the presence of gene cassette arrays in other environments. Humans are in
335 contact with animals, animal food products and other environmental sources of
336 integrons, and can both be recipients of and donors of integron-carrying bacteria or
337 genetic material (Stokes & Gillings, 2011). The flow of integrons and gene cassettes
338 between environments is a spatially and temporally variable, and may be best explained
339 as a multidimensional process. The specific events of gene exchange and directions will
340 depend on a multitude of factors, such as the encoded trait(s) and genetic composition
341 of the integron, the mobility of the genetic region it is embedded in, randomness and
342 chance dispersal, and the characteristics of the host and environmental conditions,
343 including selection of the host bacterium for integron encoded traits. Examples of the
344 many clinical and environmental sources in which the arrays are found are given in
345 Table 4.

346 Importantly, integrons have been detected in locations where a selective advantage of
347 carrying antibiotic resistance traits to pharmaceutically-produced antibiotics is not
348 expected, such as in wild animals (Power *et al.*, 2013) and in remote rural communities
349 with minimal consumption of antibiotics (Pallecchi *et al.*, 2007). An association
350 between the presence of class 1 integrons and urbanization has not been found in a
351 Patagonian island (Nardelli *et al.*, 2012). On the other hand, a recent study suggested
352 that human activities are a major source of class 1 integrons, which end up in the
353 environment (Stalder *et al.*, 2014). Power and colleagues (Power *et al.*, 2013) provided
354 evidence for the transfer of class 1 integrons from humans to captive rock wallabies.
355 These wallabies were later released, resulting in an opportunity to further spread the
356 integrons into wild populations of wallabies and their natural environments. Other
357 studies suggest that animals transmit antibiotic resistance to humans. For example,
358 although not integron-based, Harrison and collaborators (Harrison *et al.*, 2013)
359 suggested the transmission of methicillin-resistant *Staphylococcus aureus* from
360 livestock to humans.

361

362

363 **Observational bias and other limitations in studies**

364 It is emphasized that the species, geographical and environmental distribution patterns
365 of class 1 integrons examined here do not represent the actual prevalence pattern.
366 Limitations in experimental design, sampling and reporting will lead to observational
367 bias, and dominance of studies from research-oriented countries and particular clinical
368 environments in the published scientific literature. The urgency of communicating
369 clinical resistance developments creates a positive publication bias in the scientific
370 literature on the presence of integrons in such environments.

371 Large differences between countries in research focus and funding and in public health
372 monitoring and reporting systems will, moreover, necessarily lead to non-consistent
373 publication of data that are not representative for the dissemination of integrons at the
374 global scale. It is also recognized that the prevalence of particular cassette arrays are
375 expected to vary over time, so any overview of distribution patterns will be a snapshot
376 based on the available literature - with a time lag.

377 A limitation to the current study of class 1 integrons is that most studies are qualitative
378 and descriptive in nature. The experimental study design is based on the need to better
379 understand the genetic basis associated with the specific resistances observed in clinical
380 isolates of bacterial pathogens. Such studies are often done with only limited
381 consideration of the underlying population structure of the pathogen/species examined.
382 These case and isolate-focused studies do therefore not permit a quantitative analysis of
383 integrons as present in a particular population of a bacterial species or community of
384 several bacterial species. Quantitative analyses are, however, needed to enable
385 monitoring of changes in actual prevalence of integrons over time, i.e. changes in the
386 overall proportion of members of a particular bacterial population carrying a specific
387 gene cassette array.

388 Another limitation to the studies of the prevalence of integrons is the lack of sufficient
389 or complete information of the genetic composition of various bacterial communities.
390 Lack of information on the genetic composition of natural bacterial communities
391 precludes the opportunity to unambiguously track or assign directionality in the flow of
392 gene cassettes between anthropogenic and natural environments.

393

394 **General considerations**

395 Our review highlights the broad diversity of bacterial species sharing the same gene
396 cassette arrays. These species have been recovered from a wide range of environments
397 (e.g. humans; domestic, food and wild animals; and various environmental sites) located

398 in all regions of the world. This review substantiates further the observations of
399 Partridge and collaborators (Partridge *et al.*, 2009) on the broad distribution of these
400 particular sequences in class 1 integrons.

401 The current dissemination pattern of particular and sequence identical gene cassette
402 arrays suggests that opportunities exist for rapid clonal dissemination between
403 geographical locations and environments on a global scale, as well as the presence of
404 functional and effective mechanisms for horizontal gene transfer (HGT) between
405 unrelated bacterial species. It is also noteworthy that whereas the initial reports of
406 integrons focused mostly on their presence in bacteria from clinical specimens, several
407 recent studies now describe the dissemination of the same integrons in species rarely
408 associated with clinical disease. The impact of various sources of observational bias,
409 and limitations in sampling in the investigating and reporting the dissemination of
410 mobile integrons in species, environments and geographical areas remains unexplored,
411 but we consider it to be substantial.

412

413 Emergence and persistence

414 Integrons have been found in a permafrost sample (Petrova *et al.*, 2011) and in bacteria
415 isolated years before their characterization as mobile genetic elements involved in
416 antibiotic resistance (Dalsgaard *et al.*, 2000; Hedges *et al.*, 1972; Huovinen & Jacoby,
417 1991; Kazama *et al.*, 1995; Stokes & Hall, 1989). These early observations support the
418 view that integrons are not recent entities and that they have existed in bacterial
419 populations for many decades (Rowe-Magnus *et al.*, 2002). The introduction of
420 antibiotics in clinical, veterinary and agricultural settings have most likely contributed
421 to the selective amplification of new resistance-encoding gene cassettes and increased
422 the distribution and prevalence of integron-carrying bacteria with such cassettes. The
423 majority of the gene cassettes reviewed here encode resistance to antibiotics that can
424 also be produced naturally by environmental microorganisms, suggesting that particular
425 gene cassettes and integrons evolved before bacteria became exposed to
426 pharmaceutically produced antibiotics. This view is also consistent with the recent
427 finding of antibiotic resistance determinants in DNA millions of years old (D'Costa *et*
428 *al.*, 2011). There is also evidence that class 1 integrons in bacteria exposed to antibiotics
429 can capture gene cassettes from superintegrons, chromosomal integrons that contain
430 long cassette arrays (>20), where the majority of the cassettes are not usually expressed
431 due to the distance to the P_c promoter (Mazel, 2006; Rowe-Magnus *et al.*, 2002; Rowe-

432 Magnus *et al.*, 2001). Forsberg and colleagues recently also showed that soil bacteria
433 and human pathogens share the same antibiotic resistome, and that horizontal transfer
434 between both communities contributes to the resistance dissemination (Forsberg *et al.*,
435 2012). The increased prevalence of integrons is likely the outcome of selection
436 conferred by the exposure to pharmaceutically produced antimicrobials. Integrons have
437 therefore, together with other resistance-conferring mobile genetic elements, been seen
438 as xenogenetic pollutants (Gillings, 2013).

439 Integrons, similar to those found in clinical settings, have also been detected in remote
440 communities with a history of minimal exposure to antibiotics (Pallecchi *et al.*, 2007).
441 This observation emphasizes that antibiotic usage levels are not the only factor
442 responsible for the existence of integrons in current bacterial populations. Several other
443 factors might contribute to the tempospatial distribution of cassette arrays, including the
444 activity of the integrase, the phenotypic traits provided to the host by each cassette, the
445 mobility of the element where the gene cassettes are inserted, positive selection of genes
446 and genetic elements linked to the integrons, and compensatory mutations that reduce
447 the cost of integron carriage (Partridge *et al.*, 2009; Starikova *et al.*, 2012).

448 Ten different variants of the class 1 integrase, resulting from 13 different variants of the
449 gene cassette promoter P_c , have been identified. The recombination activity of each
450 integrase is inversely related to the strength of the P_c promoter (Jove *et al.*, 2010), which
451 influences the stability of the gene cassette arrays. Thus, an unresolved question is if
452 particular arrays are more widely disseminated because of a low recombination activity
453 in the class 1 integrons carrying them? Determination of the nucleotide sequence of the
454 integrase could clarify the variant present in these class 1 integrons and allow prediction
455 of the recombination activity level.

456 In bacterial species with an SOS response, the integrase gene is normally repressed,
457 which allows the host bacterium to maintain the gene cassette arrays in a steady state.
458 The SOS response is a global response to DNA damage, which controls DNA repair and
459 mutagenesis. A stress responsive regulation of integrase functions is expected to reduce
460 the biological costs of such genetic element (Guerin *et al.*, 2009). Triggering of the SOS
461 system can be observed after conjugation (Baharoglu *et al.*, 2010), natural
462 transformation (Baharoglu *et al.*, 2012) and by exposure to subinhibitory concentrations
463 of antibiotics (Baharoglu & Mazel, 2011). The activation of the SOS response leads to
464 increased transcription of the integrase gene and increased integrase activity, resulting
465 in cassette rearrangements (Baharoglu *et al.*, 2010; Guerin *et al.*, 2009). A gene cassette

466 rearrangement has also been recently observed *in vivo* after induction of the SOS
467 system, leading to emergence of a resistant *P. aeruginosa* isolate that was then quickly
468 spread among the hospital patients (Hocquet *et al.*, 2012).

469 It was recently shown that the carriage of class 1 integrases can be costly to the host, but
470 that single point mutations can inactivate the integrase restoring bacterial fitness
471 (Starikova *et al.*, 2012). The inactivation of the integrase can be an alternative way of
472 controlling fitness-reducing effects of integrase expression in bacteria in the absence of
473 SOS system, which is not present in all bacterial species. The inactivation of the
474 integrase will contribute to a higher stability of the cassette arrays embedded in an
475 integron.

476

477 Clonal (vertical) dissemination versus horizontal gene transfer

478 A key question that emerges from the observation of a wide dissemination of class 1
479 integrons and associated cassette arrays is to what extent the current pattern is a result of
480 rapid clonal amplification and spread *or* highly efficient HGT mechanisms. The rapid
481 population expansion of particular bacterial clones carrying integrons can occur both as
482 a result of that the integrons enhance the fitness of their bacterial host through the traits
483 they encode, or that integrons can hitchhike with clones that are evolutionary successful
484 for other reasons. The dispersal pattern can also results from frequent intra and
485 interspecies HGT events. The mobility of class 1 integrons through various pathways of
486 HGT has been recently reviewed (Domingues *et al.*, 2012a; Domingues *et al.*, 2012b).
487 Often both clonal expansion and HGT events act together and contribute to the
488 dissemination of resistant bacteria, as observed by Krauland and colleagues (Krauland
489 *et al.*, 2009). Some studies suggest a larger role of clonal dissemination (Kouda *et al.*,
490 2009) and others horizontal transfer (Blahna *et al.*, 2006) in such propagation.

491 On one hand, clonal dissemination of integron-carrying bacteria can explain the
492 occurrence of a particular integron in a specific bacterial species in different
493 geographical regions. For instance, the high occurrence of the *aadA2* and *bla*_{CARB-2}
494 cassettes can be explained by their presence in the pathogenic *S. enterica* serovar
495 Typhimurium DT104, which has been extensively sequenced (Antunes *et al.*, 2006;
496 Casin *et al.*, 1999; Zhao *et al.*, 2007). Increased international travel in the last decades
497 has increased the potential for dissemination of virulent clones carrying integrons. For
498 instance, class 1 integrons were detected in 11 clinical isolates of *Shigella* spp. in Spain,
499 which were associated with travel from other countries (Navia *et al.*, 2004). There is

500 also the example of integron-containing *A. baumannii* clinical isolates found in the
501 United States and the United Kingdom, which were associated with patients travelling
502 from Iraq (Turton *et al.*, 2006).

503 On the other hand, the occurrence of integrons with the same nucleotide sequence in
504 genetically unrelated species clearly identify a HGT as a causal mechanism in the
505 dissemination of integrons. The observation of conserved gene cassette compositions in
506 a high variety of bacterial species emphasizes the unusual and broad potential for
507 horizontal dissemination of gene cassettes and integrons. The outcome of such HGT
508 processes has been the global dissemination of particular resistance traits and
509 combinations within a few decades. It is noted that horizontal exchange of genetic
510 material conferring antibiotic resistance can occur independent of the genetic
511 relatedness of the involved bacterial species by conjugation, transduction or
512 transformation (Domingues *et al.*, 2012a; Domingues *et al.*, 2012c). Historically, class 1
513 integrons have been associated with Tn402-like transposons and have later been
514 incorporated in transposons like Tn21 and Tn1696 due to shared recombination
515 mechanisms (Gillings *et al.*, 2008; Partridge *et al.*, 2001; Stokes *et al.*, 2006). However,
516 the population genetic processes that have produced the current dissemination patterns
517 of integrons remain to be fully understood.

518

519 **Concluding remarks**

520 A diverse set of bacterial species shares the same gene cassettes and arrays. These host
521 species have been recovered from a wide range of environments located in all regions of
522 the world. The current distribution pattern of commonly occurring gene cassette arrays
523 suggests both i) opportunities for rapid dissemination of successful clones at a global
524 scale and ii) the existence of effective routes of HGT between geographical locations,
525 environments and unrelated bacterial species. The current integron dispersal pattern has
526 developed over the last 3 decades. Studies now also emerge describing resistance
527 encoding gene cassettes in integrons in bacterial species rarely associated with clinical
528 disease, suggesting ongoing HGT and dissemination processes in the broader
529 environment. The evolutionary significance of resistance-carrying integrons from the
530 pre-antibiotic era to current resistance patterns remains to be determined. Current data
531 suggest multidirectional flow of bacteria and their integrons between humans, animals
532 and other environments (Stokes & Gillings, 2011). Such flow has major and serious
533 implications for human health and treatment of infections. We highlight the need of

534 moving from a descriptive approach to integron dissemination to one that allow a more
535 comprehensive understanding of the factors governing the prevalence, persistence and
536 spread of integrons between and within bacterial populations.

537

538

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541

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1401 Table 1 – Examples of alternative designations of gene cassettes^a.

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Gene cassette	Other designations used ^a	
<i>aadA1a</i>	<i>ant(3')</i> -1 <i>a</i> ; <i>aad(3')</i> (9); <i>aadA1</i> ; <i>aadA</i>	1403
<i>aadA5</i>	<i>ant(3')</i> -1 <i>e</i>	1404
<i>aadB</i>	<i>ant(2')</i> -1 <i>a</i>	1405
<i>bla</i> _{CARB-2}	<i>blaP1</i> ; <i>blaP1b</i> ; <i>blaPSE-1</i>	1406
<i>dfrA1</i>	<i>dfr1</i> ; <i>dhfrI</i> ; <i>dhfrIb</i>	1407
<i>dfrA12</i>	<i>dfr12</i> ; <i>dhfrXII</i>	1408
<i>dfrA17</i>	<i>dfr17</i> ; <i>dhfrXVII</i>	1409
<i>dfrA7</i>	<i>dfrVII</i> ; <i>dhfrVII</i>	1410
<i>gcuC</i>	<i>orf</i> ; <i>orfI</i> ; <i>orfC</i> ; <i>orfX</i>	1411
<i>gcuF</i>	<i>orfF</i>	1412
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1416 ^aThese are also valid names. A lack of standards for indicating variants of gene cassettes is noted.

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1420 Table 2 – Examples of the occurrence of common gene cassette arrays in class 1 integrons in diverse bacterial species

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Gene cassette array	<i>aadB</i>	<i>dfrA7</i>	<i>aadA1a</i>	<i>aadA2</i>	<i>bla</i> _{CARB-2}	<i>dfrA1-gcuC</i>	<i>dfrA1-aadA1a</i>	<i>dfrA17-aadA5</i>	<i>dfrA12-gcuF-aadA2</i>	Total nr arrays
Species										
<i>Achromobacter xylosoxidans</i>							Traglia <i>et al.</i> , 2012			1
<i>Acidovorax defluvii</i>			Li <i>et al.</i> , 2009							1
<i>Acinetobacter baumannii</i>	Nemec <i>et al.</i> , 2004	Segal <i>et al.</i> , 2003		Gu <i>et al.</i> , 2007	Koh <i>et al.</i> , 2007	Kansakar <i>et al.</i> , 2011	Valenzuela <i>et al.</i> , 2007	Gu <i>et al.</i> , 2007		7
<i>Acinetobacter nosocomialis</i>		Xu <i>et al.</i> , 2008a			Valenzuela <i>et al.</i> , 2007		Valenzuela <i>et al.</i> , 2007			3
<i>Aeromonas allosaccharophila</i>							Moura <i>et al.</i> , 2012			1
<i>Aeromonas bestiarum</i>								Kadlec <i>et al.</i> , 2011		1
<i>Aeromonas caviae</i>		Barlow <i>et al.</i> , 2008		Carvalho <i>et al.</i> , 2012				Kadlec <i>et al.</i> , 2011		3
<i>Aeromonas eucrenophila</i>								Carvalho <i>et al.</i> , 2012		1
<i>Aeromonas hydrophila</i>		Lee <i>et al.</i> , 2008		Lukkana <i>et al.</i> , 2012		Lukkana <i>et al.</i> , 2012		Carvalho <i>et al.</i> , 2012		4
<i>Aeromonas media</i>		Moura <i>et al.</i> , 2012		Carvalho <i>et al.</i> , 2012				Moura <i>et al.</i> , 2012		3
<i>Aeromonas punctata</i>						Xu <i>et al.</i> , 2011a		FM957886 ^a		2
<i>Aeromonas</i>		L'Abée-Lund &		L'Abée-Lund &			Kadlec <i>et al.</i> ,			3

<i>salmonicida</i>	Sorum, 2001	Sorum, 2001	2011			
<i>Aeromonas sobria</i>			Kadlec <i>et al.</i> , 2011		1	
<i>Aeromonas</i> sp.			Tacao <i>et al.</i> , 2014		1	
<i>Aeromonas veronii</i>	Moura <i>et al.</i> , 2012	Ndi & Barton, 2011	Kadlec <i>et al.</i> , 2011	Carvalho <i>et al.</i> , 2012	4	
<i>Alcaligines faecalis</i>	Barlow <i>et al.</i> , 2008				1	
<i>Arcanobacterium</i> <i>pyogenes</i>		Zhao <i>et al.</i> , 2011			1	
<i>Bacillus endophyticus</i>	Li <i>et al.</i> , 2009				1	
<i>Brevundimonas</i> sp.	Yang <i>et al.</i> , 2010b				1	
<i>Buttiauxella agrestis</i>			Chen <i>et al.</i> , 2010		1	
<i>Campylobacter coli</i>		O'Halloran <i>et al.</i> , 2004			1	
<i>Campylobacter jejuni</i>		O'Halloran <i>et al.</i> , 2004			1	
<i>Citrobacter</i> <i>amalonaticus</i>	Pepperell <i>et al.</i> , 2002				1	
<i>Citrobacter braakii</i>	Mokracka <i>et al.</i> , 2012				1	
<i>Citrobacter freundii</i>	Frank <i>et al.</i> , 2007	Mokracka <i>et al.</i> , 2012	Pepperell <i>et al.</i> , 2002	Mokracka <i>et al.</i> , 2012	JN645876 ^a	Mokracka <i>et al.</i> , 2012
<i>Citrobacter koseri</i>					Mokracka <i>et al.</i> , 2012	1
<i>Citrobacter youngae</i>		Srinivasan <i>et al.</i> , 2008				1

<i>Corynebacterium glutamicum</i>			Nesvera <i>et al.</i> , 1998					1
<i>Enterobacter aerogenes</i>						Su <i>et al.</i> , 2011	Kor <i>et al.</i> , 2013	2
<i>Enterobacter cloacae</i>	Frank <i>et al.</i> , 2007	Hussein <i>et al.</i> , 2009	Dahmen <i>et al.</i> , 2010		Mokracka <i>et al.</i> , 2012	Mokracka <i>et al.</i> , 2012	Mokracka <i>et al.</i> , 2012	6
<i>Enterobacter hormaechei</i>			Mokracka <i>et al.</i> , 2011			Mokracka <i>et al.</i> , 2011	Mokracka <i>et al.</i> , 2011	3
<i>Enterobacter intermedius</i>			Bado <i>et al.</i> , 2010		Mokracka <i>et al.</i> , 2012	Pellegrini <i>et al.</i> , 2009		3
<i>Enterobacter sakazakii</i>			Mokracka <i>et al.</i> , 2012					1
<i>Enterococcus faecalis</i>			Clark <i>et al.</i> , 1999				Shi <i>et al.</i> , 2006a	2
<i>Escherichia coli</i>	Cameron <i>et al.</i> , 1986	Frank <i>et al.</i> , 2007	Lay <i>et al.</i> , 2012	Su <i>et al.</i> , 2012	Wu <i>et al.</i> , 2012	Vinue <i>et al.</i> , 2011	Wu <i>et al.</i> , 2012	Hsu <i>et al.</i> , 2006
<i>Klebsiella mobilis</i>			Koczura <i>et al.</i> , 2011			Mokracka <i>et al.</i> , 2012		2
<i>Klebsiella ornithinolytica</i>			Mokracka <i>et al.</i> , 2012				Mokracka <i>et al.</i> , 2012	Mokracka <i>et al.</i> , 2012
<i>Klebsiella oxytoca</i>	Kor <i>et al.</i> , 2013		Mokracka <i>et al.</i> , 2012	Peters <i>et al.</i> , 2001	Ahmed & Shimamoto, 2011	Mokracka <i>et al.</i> , 2012	JX560787 ^a	6
<i>Klebsiella pneumoniae</i>	Gruteke <i>et al.</i> , 2003	Frank <i>et al.</i> , 2007	Mokracka <i>et al.</i> , 2012	Gruteke <i>et al.</i> , 2003	Kor <i>et al.</i> , 2013	Wu <i>et al.</i> , 2012	Frank <i>et al.</i> , 2007	Wu <i>et al.</i> , 2012
<i>Kluyvera georgiana</i>							Rodriguez <i>et al.</i> , 2010	
<i>Kluyvera</i> sp.		Mokracka <i>et al.</i> , 2012	Mokracka <i>et al.</i> , 2012			Mokracka <i>et al.</i> , 2012		3

<i>Laribacter</i>				Feng <i>et al.</i> , 2011	Feng <i>et al.</i> , 2011			2		
<i>hongkongensis</i>										
<i>Leclercia</i>							Shin <i>et al.</i> , 2012	1		
<i>adecorboxylata</i>										
<i>Morganella morganii</i>	Mahrouki <i>et al.</i> , 2012	Hussein <i>et al.</i> , 2009	Barlow <i>et al.</i> , 2008	Kor <i>et al.</i> , 2013	Kor <i>et al.</i> , 2013	Kor <i>et al.</i> , 2013		6		
<i>Ochrobactrum</i> sp.							Li <i>et al.</i> , 2010	1		
<i>Pantoea agglomerans</i>		Aibinu <i>et al.</i> , 2012	Barlow <i>et al.</i> , 2008					2		
<i>Pantoea</i> sp.		Mokracka <i>et al.</i> , 2012				Mokracka <i>et al.</i> , 2012		2		
<i>Proteus mirabilis</i>	JX494728*	EU860402*	Gionechetti <i>et al.</i> , 2008	Falcone <i>et al.</i> , 2010	Boyd <i>et al.</i> , 2008	Gionechetti <i>et al.</i> , 2008	Wu <i>et al.</i> , 2012	7		
<i>Proteus vulgaris</i>		Hussein <i>et al.</i> , 2009	Hussein <i>et al.</i> , 2009			Ozgumus <i>et al.</i> , 2009		3		
<i>Providencia</i>				Xu <i>et al.</i> , 2011a				1		
<i>alcalifaciens</i>										
<i>Pseudomonas</i>	Ruiz-Martinez <i>et al.</i> , 2011		Gu <i>et al.</i> , 2007	HQ832476*		Xu <i>et al.</i> , 2009	Gu <i>et al.</i> , 2007	5		
<i>aeruginosa</i>										
<i>Pseudomonas putida</i>			Wu <i>et al.</i> , 2012					1		
<i>Pseudomonas</i> sp.		Rosser & Young, 1999						1		
<i>Raoultella planticola</i>							Tseng <i>et al.</i> , 2014	1		
<i>Riemerella</i>			Zheng <i>et al.</i> , 2012	Sun <i>et al.</i> , 2012				2		
<i>anatipestifer</i>										
<i>Salmonella enterica</i>	Zhao <i>et al.</i> , 2007	Tamang <i>et al.</i> , 2007	Ranjbar <i>et al.</i> , 2011	Kim <i>et al.</i> , 2011	Havlickova <i>et al.</i> , 2009	Lee <i>et al.</i> , 2009	Perez-Moreno <i>et al.</i> , 2013	Perez-Moreno <i>et al.</i> , 2013	Antunes <i>et al.</i> , 2006	9

<i>Serratia liquefaciens</i>		Mokracka <i>et al.</i> , 2012					1	
<i>Serratia marcescens</i>	Peng <i>et al.</i> , 2007		Peng <i>et al.</i> , 2007	Peng <i>et al.</i> , 2007	Mokracka <i>et al.</i> , 2012	Peng <i>et al.</i> , 2007	5	
<i>Serratia odorifera</i>		Mokracka <i>et al.</i> , 2012			Mokracka <i>et al.</i> , 2012		2	
<i>Serratia</i> sp.					Chakraborty <i>et al.</i> , 2013		1	
<i>Shigella boydii</i>	Frank <i>et al.</i> , 2007						1	
<i>Shigella dysenteriae</i>	Frank <i>et al.</i> , 2007					Iversen <i>et al.</i> , 2003	2	
<i>Shigella flexneri</i>	Frank <i>et al.</i> , 2007	Navia <i>et al.</i> , 2004	Iversen <i>et al.</i> , 2003		Navia <i>et al.</i> , 2004	Pan <i>et al.</i> , 2006	Iversen <i>et al.</i> , 2003	6
<i>Shigella sonnei</i>		Nogradi <i>et al.</i> , 2013	Pan <i>et al.</i> , 2006		Navia <i>et al.</i> , 2004	Pan <i>et al.</i> , 2006	Iversen <i>et al.</i> , 2003	5
<i>Sphingobacterium</i> sp.						Li <i>et al.</i> , 2010		1
<i>Staphylococcus aureus</i>		KF687971*	Xu <i>et al.</i> , 2007			Xu <i>et al.</i> , 2011b	Xu <i>et al.</i> , 2011b	4
<i>Staphylococcus</i> CoN							Shi <i>et al.</i> , 2006a	1
<i>Staphylococcus</i> <i>epidermidis</i>			Xu <i>et al.</i> , 2008b			Xu <i>et al.</i> , 2008b	Xu <i>et al.</i> , 2008b	3
<i>Staphylococcus</i> <i>haemolyticus</i>							Xu <i>et al.</i> , 2008b	1
<i>Staphylococcus</i> <i>hominis</i>						Xu <i>et al.</i> , 2008b	Xu <i>et al.</i> , 2008b	2
<i>Staphylococcus</i> <i>warneri</i>							Xu <i>et al.</i> , 2008b	1

<i>Stenotrophomonas</i>	JX560784*			Xu <i>et al.</i> , 2011a	Hu <i>et al.</i> , 2011	Hu <i>et al.</i> , 2011	4		
<i>maltophilia</i>									
<i>Stenotrophomonas</i> sp.		Yang <i>et al.</i> , 2010b					1		
<i>Streptococcus</i> spp.			Shi <i>et al.</i> , 2006a			Shi <i>et al.</i> , 2006a	2		
<i>Vibrio alginolyticus</i>			Taviani <i>et al.</i> , 2008				1		
<i>Vibrio cholerae</i>	Dalsgaard <i>et al.</i> , 2000	Ceccarelli <i>et al.</i> , 2006a	Dalsgaard <i>et al.</i> , 2000	Dalsgaard <i>et al.</i> , 2000	Thungapath ra <i>et al.</i> , 2002		5		
<i>Vibrio fluvialis</i>		Rajpara <i>et al.</i> , 2009			Srinivasan <i>et al.</i> , 2006		2		
<i>Xantomonas oryzae</i>		FJ501978*					1		
Total nr species	11	15	39	31	6	12	22	26	37

* GenBank accession number

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1431 Table 3 – Examples of the geographical distribution of some of the most common gene cassette arrays embedded in class 1 integrons

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Country	Gene cassette array	<i>aadB</i>	<i>dfrA7</i>	<i>aadA1a</i>	<i>aadA2</i>	<i>bla</i> _{CARB-2}	<i>dfrA1-gcuC</i>	<i>dfrA1-aadA1 a</i>	<i>dfrA17-aadA5</i>	<i>dfrA12-gcuF-aadA2</i>	Total n ^{er} arrays
Albania				Falbo <i>et al.</i> , 1999							1
Angola						Ceccarelli <i>et al.</i> , 2006b					1
Argentina				Di Conza <i>et al.</i> , 2005				Traglia <i>et al.</i> , 2012			2
Australia	Jones <i>et al.</i> , 2005	Bailey <i>et al.</i> , 2010	Barlow <i>et al.</i> , 2008	Bailey <i>et al.</i> , 2010	Valenzuela <i>et al.</i> , 2007		Bailey <i>et al.</i> , 2010	Sidjabat <i>et al.</i> , 2006		Power <i>et al.</i> , 2013	8
Austria				Morabito <i>et al.</i> , 2002							1
Belgium	Huys <i>et al.</i> , 2005						Plante <i>et al.</i> , 2003				2
Bolivia		Pallecchi <i>et al.</i> , 2007					Pallecchi <i>et al.</i> , 2007	Pallecchi <i>et al.</i> , 2007			3
Brazil	DQ139277*			Peirano <i>et al.</i> , 2006	Sa <i>et al.</i> , 2010					Peirano <i>et al.</i> , 2006	4
Canada	Allen & Poppe, 2002		Pepperell <i>et al.</i> , 2002	Pepperell <i>et al.</i> , 2002	Ng <i>et al.</i> , 1999	Xu <i>et al.</i> , 2011a	Xu <i>et al.</i> , 2011a		Wu <i>et al.</i> , 2011	Wu <i>et al.</i> , 2011	8
Central African Republic		Frank <i>et al.</i> , 2007					Frank <i>et al.</i> , 2007	Frank <i>et al.</i> , 2007			3
Chile		Lapierre <i>et al.</i> , 2008					Lapierre <i>et al.</i> , 2008				2

China	Xu <i>et al.</i> , 2009	Su <i>et al.</i> , 2012	Lu <i>et al.</i> , 2010	Su <i>et al.</i> , 2012	Yang <i>et al.</i> , 2010a	Feng <i>et al.</i> , 2011	Su <i>et al.</i> , 2012	Su <i>et al.</i> , 2012	Su <i>et al.</i> , 2012	9
Colombia								GU304661*		1
Czech Republic		Dolejska <i>et al.</i> , 2008		Havlickova <i>et al.</i> , 2009		Dolejska <i>et al.</i> , 2008	Dolejska <i>et al.</i> , 2009			4
Denmark	Sandvang & Krauland <i>et al.</i> , 2000	Krauland <i>et al.</i> , 2009	Sandvang & Aarestrup, 1998	Sandvang <i>et al.</i> , 1998	Sandvang <i>et al.</i> , 2000			Krauland <i>et al.</i> , 2009		6
Egypt		Ahmed <i>et al.</i> , 2009b	Ahmed & Shimamoto, 2011			Ahmed <i>et al.</i> , 2009b	Ahmed <i>et al.</i> , 2009b	Ahmed <i>et al.</i> , 2009b		5
Ethiopia		Molla <i>et al.</i> , 2007	Molla <i>et al.</i> , 2007			Molla <i>et al.</i> , 2007		Molla <i>et al.</i> , 2007		4
Finland								Heikkila <i>et al.</i> , 1993		1
France	Huys <i>et al.</i> , 2005		Vinue <i>et al.</i> , 2011	Casin <i>et al.</i> , 1999	Poirel <i>et al.</i> , 1999		Vinue <i>et al.</i> , 2011	Vinue <i>et al.</i> , 2011	Vinue <i>et al.</i> , 2011	7
Germany	Rodriguez <i>et al.</i> , 2009	Blahna <i>et al.</i> , 2006	Kadlec & Schwarz, 2008	Heuer & Smalla, 2007	Rodriguez <i>et al.</i> , 2009		Kadlec & Schwarz, 2008	Kadlec & Schwarz, 2008	Kadlec & Schwarz, 2008	8
Ghana		Labar <i>et al.</i> , 2012	Labar <i>et al.</i> , 2012				Labar <i>et al.</i> , 2012	Labar <i>et al.</i> , 2012		4
Gibraltar		Ridley & Threlfall, 1998								1
Hungary	Libisch <i>et al.</i> , 2009		Nogrady <i>et al.</i> , 2005	Nogrady <i>et al.</i> , 2005	Nogrady <i>et al.</i> , 2005		Nogrady <i>et al.</i> , 2005	Nogrady <i>et al.</i> , 2005		6
India		Ploy <i>et al.</i> , 2003	Thungapathra <i>et al.</i> , 2002	Shi <i>et al.</i> , 2006b	Shi <i>et al.</i> , 2006b		HE653235*	Pazhani <i>et al.</i> , 2011	FM179328*	7
Indonesia							Waturangi <i>et al.</i> , 2003			2
Iran	Shahcheraghi <i>et al.</i> , 2010	Najibi <i>et al.</i> , 2012	Ranjbar <i>et al.</i> , 2011	Adabi <i>et al.</i> , 2009	HQ132377*	Srinivasan <i>et al.</i> , 2006	Ranjbar <i>et al.</i> , 2011		Najibi <i>et al.</i> , 2012	8

Ireland		Karczmarczyk <i>et al.</i> , 2011	Karczmarczyk <i>et al.</i> , 2011	O'Halloran <i>et al.</i> , 2004	Murphy <i>et al.</i> , 2007	Karczmarczyk <i>et al.</i> , 2011	Karczmarczyk <i>et al.</i> , 2011	Karczmarczyk <i>et al.</i> , 2011	7
Italy	Huys <i>et al.</i> , 2005		Falbo <i>et al.</i> , 1999	Falcone <i>et al.</i> , 2010	GU987051 ^a	Gionechetti <i>et al.</i> , 2008	Gionechetti <i>et al.</i> , 2008	Morabito <i>et al.</i> , 2002	7
Japan		Ahmed <i>et al.</i> , 2009a	Ahmed <i>et al.</i> , 2009a	Yokoyama <i>et al.</i> , 2007	Ahmed <i>et al.</i> , 2005	Ahmed <i>et al.</i> , 2007	Ahmed <i>et al.</i> , 2007	Kumai <i>et al.</i> , 2005	8
Jordan		Al-Sanouri <i>et al.</i> , 2008							1
Kenya		Kikuvi <i>et al.</i> , 2007	Kiiru <i>et al.</i> , 2013			Kikuvi <i>et al.</i> , 2007	Kiiru <i>et al.</i> , 2013	Kiiru <i>et al.</i> , 2013	5
Korea	Kang <i>et al.</i> , 2010	Lee <i>et al.</i> , 2004	Kang <i>et al.</i> , 2005	Yu <i>et al.</i> , 2003	Lee & Lee, 2007	Kim <i>et al.</i> , 2011	Kang <i>et al.</i> , 2005	Kang <i>et al.</i> , 2005	8
Lebanon		El-Najjar <i>et al.</i> , 2010				El-Najjar <i>et al.</i> , 2010	El-Najjar <i>et al.</i> , 2010		3
Lithuania	JF412714*		Povilonis <i>et al.</i> , 2010			Povilonis <i>et al.</i> , 2010	Povilonis <i>et al.</i> , 2010	Povilonis <i>et al.</i> , 2010	4
Madagascar			Rakotonirina <i>et al.</i> , 2013	Rakotonirina <i>et al.</i> , 2013			Rakotonirina <i>et al.</i> , 2013		3
Malaysia	Kor <i>et al.</i> , 2013	Kor <i>et al.</i> , 2013	Kor <i>et al.</i> , 2013	Mukherjee & Chakraborty, 2013	Kor <i>et al.</i> , 2013	Kor <i>et al.</i> , 2013	Kor <i>et al.</i> , 2013	Kor <i>et al.</i> , 2013	9
Mexico				Perez-Valdespino <i>et al.</i> , 2009			Wiesner <i>et al.</i> , 2009	Wiesner <i>et al.</i> , 2009	3
Mozambique		Ceccarelli <i>et al.</i> , 2006a	Taviani <i>et al.</i> , 2008	Taviani <i>et al.</i> , 2008					3
Nepal		Tamang <i>et al.</i> , 2007							1
Nigeria		Labar <i>et al.</i> , 2012	Labar <i>et al.</i> , 2012	Labar <i>et al.</i> , 2012		Labar <i>et al.</i> , 2012	Labar <i>et al.</i> , 2012	Labar <i>et al.</i> , 2012	6

Norway		Heir <i>et al.</i> , 2004	Sunde, 2005	Lindstedt <i>et al.</i> , 2003		Sunde, 2005	Lindstedt <i>et al.</i> , 2003	Lindstedt <i>et al.</i> , 2003	6
Pakistan		Holt <i>et al.</i> , 2007							1
Palestine territory	Hussein <i>et al.</i> , 2009		Hussein <i>et al.</i> , 2009	Hussein <i>et al.</i> , 2009			Hussein <i>et al.</i> , 2009	Hussein <i>et al.</i> , 2009	6
Philippines							Krauland <i>et al.</i> , 2009		1
Poland		Mokracka <i>et al.</i> , 2012	Mokracka <i>et al.</i> , 2012			Mokracka <i>et al.</i> , 2012	Mokracka <i>et al.</i> , 2012	Mokracka <i>et al.</i> , 2012	5
Portugal	Ferreira da Silva <i>et al.</i> , EU860402* 2007		Antunes <i>et al.</i> , 2006	Moura <i>et al.</i> , 2007	Antunes <i>et al.</i> , 2006	Moura <i>et al.</i> , 2007	Antunes <i>et al.</i> , 2006	Carvalho <i>et al.</i> , 2012	8
Russia	GQ924772*		GQ924774*	Petrova <i>et al.</i> , 2011	HQ832476*	GQ924770*	GQ896490*	Egorova <i>et al.</i> , 2007	7
Rwanda		Ceccarelli <i>et al.</i> , 2006a							1
Scotland			L'Abée-Lund & Sorum, 2001						1
Senegal		Gassama <i>et al.</i> , Gassama <i>et al.</i> , 2004				Sow <i>et al.</i> , 2007			3
Singapore				Koh <i>et al.</i> , 2007			FM957886*		2
Slovakia		Majtan <i>et al.</i> , 2007	Majtan <i>et al.</i> , 2007	Majtan <i>et al.</i> , 2007		Majtan <i>et al.</i> , 2007		Majtan <i>et al.</i> , 2007	5
Slovenia		Ridley & Threlfall, 1998							1
Somalia		Ceccarelli <i>et al.</i> , 2006a							1
South Africa		Krauland <i>et al.</i> , 2009	Ridley & Threlfall, 1998	Dalsgaard <i>et al.</i> , 2001			Krauland <i>et al.</i> , 2009		4
Spain	Huys <i>et al.</i> , 2005	Perez-Moreno <i>et al.</i> , 2013	Rodriguez <i>et al.</i> , 2006	Perez-Moreno <i>et al.</i> , 2013	Guerra <i>et al.</i> , 2000	Perez-Moreno <i>et al.</i> , 2013	Perez-Moreno <i>et al.</i> , 2013	Perez-Moreno <i>et al.</i> , 2013	8

Sri Lanka		Sundstrom <i>et al.</i> , 1987							1
Sudan		Ibrahim <i>et al.</i> , 2013			Ibrahim <i>et al.</i> , 2013	Ibrahim <i>et al.</i> , 2013	Ibrahim <i>et al.</i> , 2013		4
Swaziland		Ceccarelli <i>et al.</i> , 2006a							1
Sweden	Tennhammar- Ekman & Skold, 1979	Grape <i>et al.</i> , 2005	Grape <i>et al.</i> , 2005		Grape <i>et al.</i> , 2005	Grape <i>et al.</i> , 2005	Grape <i>et al.</i> , 2005		6
Switzerland		Cocchi <i>et al.</i> , 2007			Cocchi <i>et al.</i> , 2007	Cocchi <i>et al.</i> , 2007			2
Taiwan	Peng <i>et al.</i> , 2007 2009	Yang <i>et al.</i> , 2009	Hsu <i>et al.</i> , 2006	Hsu <i>et al.</i> , 2006	Lee <i>et al.</i> , 2009	Chang <i>et al.</i> , 2000	Hsu <i>et al.</i> , 2006	Hsu <i>et al.</i> , 2006	8
Thailand	Dalsgaard <i>et al.</i> , 2000		Lay <i>et al.</i> , 2012	Dalsgaard <i>et al.</i> , 2000	Dalsgaard <i>et al.</i> , 2000			Lukkana <i>et al.</i> , 2012	6
The Netherlands	Huys <i>et al.</i> , 2005	Vo <i>et al.</i> , 2007	van Essen-Zandbergen <i>et al.</i> , 2003	Gruteke <i>et al.</i> , 2007	van Essen-Zandbergen <i>et al.</i> , 2007	Vo <i>et al.</i> , 2007	van Essen-Zandbergen <i>et al.</i> , 2007	van Essen-Zandbergen <i>et al.</i> , 2007	8
Trinidad			Ridley & Threlfall, 1998						1
Tunisia	Mahrouki <i>et al.</i> , 2012	Dahmen <i>et al.</i> , 2010	Ben Sallem <i>et al.</i> , 2012	Dahmen <i>et al.</i> , 2010	Soufi <i>et al.</i> , 2012	Mahrouki <i>et al.</i> , 2012	Dahmen <i>et al.</i> , 2010	Dahmen <i>et al.</i> , 2010	9
Turkey		Ozgumus <i>et al.</i> , 2009	Sandalli <i>et al.</i> , 2010	Sandalli <i>et al.</i> , 2010			Ozgumus <i>et al.</i> , 2009	Sandalli <i>et al.</i> , 2010	6
Uganda		Krauland <i>et al.</i> , 2009	Guerra <i>et al.</i> , 2006						2
United Kingdom	Turton <i>et al.</i> , 2006	Rosser & Young, 1999	Rosser & Young, 1999	Corkill <i>et al.</i> , 2005		Rosser & Young, 1999		Rosser & Young, 1999	6
United States	Turton <i>et al.</i> , 2006	Ajiboye <i>et al.</i> , 2006	Li <i>et al.</i> , 2006	Zhao <i>et al.</i> , 2007	Zhao <i>et al.</i> , 2007	Khan <i>et al.</i> , 2006	Ajiboye <i>et al.</i> , 2009	Solberg <i>et al.</i> , 2006	9
				2009		2003		Solberg <i>et al.</i> , 2006	

Uruguay	Bado <i>et al.</i> , 2010		Bado <i>et al.</i> , 2010
Vietnam	Ploy <i>et al.</i> , 2003	Ploy <i>et al.</i> , 2003	Vo <i>et al.</i> , 2010
			Van <i>et al.</i> , 2007
			DQ238103*
			Van <i>et al.</i> , 2007
			Van <i>et al.</i> , 2007
Zimbabwe	Ceccarelli <i>et al.</i> , 2006a		
Total n^{er} countries	24	37	54
			38
			27
			11
			44
			38
			42

* GenBank accession number

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1439 Table 4 – Examples of the source of bacteria containing common gene cassette arrays embedded in class 1 integrons

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Gene cassette array	<i>aadB</i>	<i>dfrA7</i>	<i>aadA1a</i>	<i>aadA2</i>	<i>bla_{CARB-2}</i>	<i>dfrA1-gcuC</i>	<i>dfrA1-aadA1a</i>	<i>dfrA17-aadA5</i>	<i>dfrA12-gcuF-aadA2</i>	Total n^{er} arrays
Source										
Human										
Clinical		Solberg <i>et al.</i> , 2006	Nogradi <i>et al.</i> , 2005	Kazama <i>et al.</i> , 1995	Kor <i>et al.</i> , 2013	Thungapathra <i>et al.</i> , 2002	Nogradi <i>et al.</i> , 2005	White <i>et al.</i> , 2000	Heikkila <i>et al.</i> , 1993	8
Healthy		Kang <i>et al.</i> , 2005	Labar <i>et al.</i> , 2012	Labar <i>et al.</i> , 2012	Molla <i>et al.</i> , 2007		Labar <i>et al.</i> , 2012	Kang <i>et al.</i> , 2005	Kang <i>et al.</i> , 2005	7
Animal										
Domestic	Vo <i>et al.</i> , 2007	Vo <i>et al.</i> , 2007	Kadlec & Schwarz, 2008		Rodriguez <i>et al.</i> , 2009		Kadlec <i>et al.</i> , 2011	Cocchi <i>et al.</i> , 2007	Kadlec & Schwarz, 2008	7
Food-producing	Zhao <i>et al.</i> , 2005	Kikuvi <i>et al.</i> , 2007	Zhao <i>et al.</i> , 2005	O'Halloran <i>et al.</i> , 2004	Khemtong & Chuanchuen, 2008	Du <i>et al.</i> , 2005	Kadlec <i>et al.</i> , 2011	Dotto <i>et al.</i> , 2014	Ahmed <i>et al.</i> , 2009b	9
Food product		Ahmed <i>et al.</i> , 2009a	Sunde, 2005	Dalsgaard <i>et al.</i> , 2000	Zhao <i>et al.</i> , 2003	Khan <i>et al.</i> , 2006	Antunes <i>et al.</i> , 2006	Machado <i>et al.</i> , 2008	Antunes <i>et al.</i> , 2006	8
Wild	Gionechetti <i>et al.</i> , 2008		Gionechetti <i>et al.</i> , 2008		Havlickova <i>et al.</i> , 2009		Goncalves <i>et al.</i> , 2013	Dolejska <i>et al.</i> , 2009	Caleja <i>et al.</i> , 2011	6
Zoo				Ahmed <i>et al.</i> , 2007			Ahmed <i>et al.</i> , 2007	Ahmed <i>et al.</i> , 2007		3
Water										
Lake			Yang <i>et al.</i> , 2010b							1

River	Su <i>et al.</i> , 2012	Xu <i>et al.</i> , 2011a	Su <i>et al.</i> , 2012	Ceccarelli <i>et al.</i> , 2006b	Ahmed <i>et al.</i> , 2005	Su <i>et al.</i> , 2012	Su <i>et al.</i> , 2012	Su <i>et al.</i> , 2012	8
Sewage				Guerra <i>et al.</i> , 2000					1
Spring						Ozgumus <i>et al.</i> , 2007			1
Tap						Ozgumus <i>et al.</i> , 2007			1
Wastewater	Ferreira da Silva <i>et al.</i> , 2007	Tennstedt <i>et al.</i> , 2003	Moura <i>et al.</i> , 2012	Taviani <i>et al.</i> , 2008	Taviani <i>et al.</i> , 2008	Han <i>et al.</i> , 2012	Moura <i>et al.</i> , 2012	Mokracka <i>et al.</i> , 2012	8
<hr/>									
Soil									
Dairy farm			Srinivasan <i>et al.</i> , 2008				Srinivasan <i>et al.</i> , 2008		2
Manure	Heuer <i>et al.</i> , 2012		Binh <i>et al.</i> , 2009	Heuer & Smalla, 2007					3
<hr/>									
Other									
Hospital inanimate surface	JX560784*		Xu <i>et al.</i> , 2007		GU731078*		Sidjabat <i>et al.</i> , 2006		4
Permafrost			Petrova <i>et al.</i> , 2011						1
Remote community		Pallecchi <i>et al.</i> , 2007			Pallecchi <i>et al.</i> , 2007	Pallecchi <i>et al.</i> , 2007			3
Total n^{er} sources	7	8	10	11	9	4	12	12	9

* GenBank accession number

1443 **Figure 1.** Schematic structure of a classic class 1 integron. 5'-CS is the 5' conserved segment of the integron and 3'-CS the 3' conserved segment
1444 of the integron. The 5'-CS includes the class 1 integrase gene, *intI1*; the gene cassettes promoters, P_c and P2; and the integron-associated
1445 recombination site, *attI1*. The 3'-CS includes the truncated version of a quaternary ammonium resistance gene, *qacEΔI*; the sulphonamide
1446 resistance gene, *sulI*; and an open reading frame, *orf5*. The variable region contains the gene cassettes, GC; and the recombination site of the
1447 gene cassette, *attC*.

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