

1 **Measurements of body fat is associated with markers of inflammation,**
2 **insulin resistance and lipid levels in both overweight and in lean, healthy**
3 **subjects**

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26 **ABSTRACT**

27 **BACKGROUND AND AIMS:** Low-grade inflammation is associated with fat mass in
28 overweight. Whether this association exists in lean persons is unknown.

29 Aimes were to investigate associations between anthropometric measures of fat distribution
30 and fat mass (% and kg) assessed by bioelectrical impedance analysis (BIA). Furthermore we
31 wanted to investigate the relationship between fat mass and markers of insulin resistance,
32 inflammation, and lipids in healthy subjects in different BMI categories.

33 **METHODS:** We compared 47 healthy overweight adults (BMI 26-40kg/m²) and 40 lean
34 (BMI 17-25kg/m²) matched for age and sex. Waist- and hip circumferences, waist-to-hip
35 ratio, waist-to-height ratio and triceps skinfold were used to evaluate fat distribution. BIA was
36 used to estimate fat mass (% and kg). Markers of insulin resistance, lipids, inflammation and
37 adipokines were measured.

38 **RESULTS:** Hip circumference was associated ($P<0.01$) with BIA-assessed fat mass (%) in
39 both groups (lean: regression coefficient B=0.4; overweight: B=0.5). An increase in hip
40 circumference in all tertiles was associated with higher plasma levels of leptin, CRP and C-
41 peptide in both groups.

42 **CONCLUSIONS:** Fat mass may play a role in low-grade inflammation also in subjects
43 within the normal range of BMI. Hip circumference may be a surrogate measure for fat mass
44 in subjects in different BMI categories, and may be useful for identification of people with
45 risk of developing overweight-related chronic diseases.

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47 **Keywords:** fat mass, body composition, anthropometry, bioelectric impedance, inflammation

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55 INTRODUCTION

56 The prevalence of overweight and obesity has increased dramatically worldwide (1).
57 Frequently associated health risks are insulin resistance, elevated blood pressure and
58 hypercholesterolemia, which may lead to type 2 diabetes and cardiovascular disease (2). The
59 most important determinant of these problems is not the increased body mass per se, but
60 rather the total amount of fat, its distribution in the body and metabolic factors that are related
61 to fat tissue mass (3). Fat tissue is an active endocrine organ releasing adipokines (leptin,
62 adiponectin, resistin) and inflammatory factors, e.g, interleukin (IL) –6 (4). These mediators
63 modify carbohydrate- and lipid metabolism and contribute to insulin resistance,
64 hyperlipidemia and inflammatory processes (5). It is well known that inflammatory markers
65 are associated with fat mass in overweight and obese subjects (6), but this relation between fat
66 mass and inflammatory markers in lean subjects is not well documented (7).

67 Several methods are used to measure the amount of fat in adults. One of the most
68 accurate methods is Dual-energy X-ray absorptiometry (DXA) (7), but measuring fat this way
69 is costly and not readily available in clinical practice. Bioelectrical impedance analysis (BIA)
70 is more available and widely used outside hospitals (8), and an objective, quick and non-
71 invasive method for assessment of fat and fat free mass (9, 10). Validation studies of BIA
72 against DXA showed that BIA is an adequate tool for prediction of fat (%) in healthy
73 populations (11). The most common population-level measure is probably estimation of body
74 mass index (BMI) (12). Whether BMI is a good marker to define obesity and health status is
75 debated (13). Studies have shown that BMI fails to differentiate between elevated body fat
76 and increased lean mass, especially in subjects with a BMI < 30 kg/m², a frequent cut-off for
77 obesity (12). Other anthropometric measures, such as waist circumference, hip circumference,
78 waist-to-hip ratio, waist-to-height ratio and triceps skinfold, are often used to determine fat
79 distribution (13, 14). Like BMI all these measures are just proxies of fat mass, but may predict

80 adverse outcomes (14). The INTERHEART Study showed that increasing waist-to-hip ratio
81 was a predictor of myocardial infarction in subjects with BMI < 20 kg/m², subjects with
82 recommended weight (BMI 20-25 kg/m²), as well as in overweight and obese subjects (BMI
83 > 25 kg/m²) (15). Thus in further studies of the role of adipokines and inflammation in the
84 development of metabolic disorders it will be of interest to investigate if fat mass estimated by
85 anthropometric measures can predict levels of inflammatory markers not only in overweight,
86 but also in lean subjects. Our primary study aim was therefore to determine if any of the
87 frequently used anthropometric measures of fat mass (BMI, waist circumference, waist-to-hip
88 ratio, waist-to-height ratio and triceps skinfold) were associated with BIA-measured fat mass.
89 Furthermore we wanted to investigate the relationship between the anthropometric measure
90 with the strongest correlation with BIA, and adipokines, inflammatory markers, markers of
91 insulin resistance and lipids among healthy subjects in different BMI categories.

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106 **MATERIALS AND METHODS**

107 **Subjects**

108 The study population included 47 overweight and 40 lean healthy adult volunteers
109 (M:59/F:28). The overweight group consisted of subjects available for baseline analysis in a
110 contemporary intervention trial performed in 2009. They were approached through mass
111 media and selected in accordance with the following inclusion criteria: waist circumference >
112 94 cm (men), > 80 cm (women), and BMI 26 - 40 kg/m². Exclusion criteria were type 2
113 diabetes, kidney, liver, gall bladder, coronary, endocrine or rheumatoid disease, any
114 malignancy the last 5 years, hypertension ($\geq 160/100$ mmHg), pregnancy and lactation.
115 Regular use of anti-inflammatory, lipid lowering and antihypertensive medication was not
116 permitted. In 2010, a reference group of lean subjects was recruited in the same way as the
117 overweight subjects. Inclusion criteria were: waist circumference ≤ 94 cm (men), ≤ 80 cm
118 (women), BMI 17-25 kg/m² and age 18-70 years. Exclusion criteria were the same as for the
119 overweight group. The study groups were matched on age and sex. All subjects were
120 instructed to refrain from vigorous physical activity and alcohol the day before the study visit.
121 The study protocol complied with the principles laid down in the Declaration of Helsinki, and
122 approved by the Regional Committee for Medical and Health Research Ethics. Written
123 informed consent was obtained from all participants.

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125 **Laboratory methods**

126 Venous blood samples were collected after an overnight fast (≥ 12 hours), between 8.00-10.00
127 a.m. Serum was obtained from silica gel tubes (Becton Dickinson vacutainer, Plymouth, Great
128 Britain) and kept on ice, centrifuged (1500 g for 12 minutes), aliquoted and stored at -80°C
129 until further analyses (inflammatory markers), or kept in room temperature (for standard
130 clinical chemistry) for at least 30 minutes, until centrifugation at 1500 g for 12 minutes and

131 immediately prepared for subsequent analysis. Plasma was obtained from EDTA tubes
132 (Becton Dickinson), kept on ice and centrifuged (2000 g, 4°C, 10 minutes) within 15 minutes.
133 Plasma samples were aliquoted and stored at -80 °C until further analysis.

134 Serum leptin, serum adiponectin, serum resistin, plasma IL-6, and plasma insulin-like
135 growth factor-1 (IGF-1) levels were measured by enzyme immunoassays from R&D Systems
136 (Minneapolis, USA) according the manufacturer's instructions. All analyses were performed in
137 duplicates. The coefficients of variation for intra-assay and inter-assay variability were <5%
138 and <10%, respectively, for all analyses. Standard blood chemistry and lipid parameters were
139 measured in serum or in EDTA plasma at Fürst Medical Laboratory using routine methods
140 (Oslo, Norway).

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142 **Assessment of fat mass**

143 Subjects wore light clothing and no shoes. Two trained persons performed all measurements,
144 which were performed once, except for triceps skinfold (TSF), which was measured three
145 times. Height was measured by a wall-mounted stadiometer to the nearest 0.1 cm. Weight was
146 measured by a Tanita scale (BC-418 MA, Tanita Corp., Tokyo, Japan) to the nearest 0.1 kg. A
147 correction factor of -1 kg was used to adjust for the weight of light clothing before BMI was
148 calculated. Waist- and hip circumferences were measured by a standard, non-stretch tape to
149 the nearest 0.1 cm while standing in a relaxed position with normal respiration. Waist
150 circumference was measured at a point midway between the iliac crest and the lower rib
151 margin. Hip circumference was measured as the maximum circumference of the posterior
152 buttocks and the anterior symphysis. The waist-hip ratio was calculated as waist
153 circumference/hip circumference and the waist-height ratio was calculated as waist
154 circumference/height. TSF was measured by using a Harpenden Caliper and a standard, non-
155 stretch tape on the non-dominant arm. The midpoint of the arm was measured, with the

156 measuring tape between the shoulder (acromion) and the elbow (olecranon) while the person
157 was bending the arm 90 degrees. TSF and the mid-upper-arm circumference (MUAC) were
158 measured at this midpoint. The mid-upper-arm muscle circumference (MUAMC) was
159 calculated with the equation $MUAC - (\pi \times (TSF/10)) = MUAMC \text{ (cm)}$ (2).

160 Body composition was estimated using the single frequency bioimpedance analyzer
161 Tanita scale, operating at 50 kHz, with eight-point contact electrodes (16). The electrode
162 arrangement in the system allows separate measurements for each arm and leg, the trunk, and
163 whole body. Fat mass (% and kg) were calculated from the measured resistance values,
164 height, body weight, sex, age, and standard body type (defined in the producer`s manual as
165 less than ten hour of exercise per week). Measurements were performed with the subjects
166 standing barefoot on the platform with arms slightly apart from the body.

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168 **Statistical analysis**

169 Normality distribution was assessed by looking at the QQ-plots and the distribution of the
170 histograms of the variables. Descriptive statistics were used. Independent samples t-test was
171 used for comparison between groups. Univariate linear regression analyses were applied to
172 quantify the relationship between BIA- and anthropometric measurements of body fat.
173 Variables with *P*-values < 0. 2 were included in the multivariate model. A stepwise model
174 reduction procedure was applied, where the F-ratio test was used. In this test we step-by step
175 eliminated the non-significant variables from the multivariate model. This was done to
176 compare the results with and without the non-significant variables. The reduction (elimination
177 of non-significant variables) was done until it was not possible to reduce the model any
178 further. Although the groups were matched for age and sex, we adjusted for these variables to
179 correct any mistakes done in the matching procedure. In order to analyse insulin resistance
180 markers, lipids and inflammatory markers concentration with respect to body fat, hip

181 circumference, BIA measures of fat percent and BMI were divided into tertiles and analyzed
182 with ANOVA. Sample size calculations were not performed because of the descriptive
183 design. Statistical significance was set as $P < 0.05$. The PASW 18 was used for all statistical
184 analyses (SPSS Inc., Chicago, IL).

185

186 **RESULTS**

187 **Subjects**

188 Forty-seven (33 men and 14 women) overweight (BMI 25-40 kg/m²) whereof 25 were obese
189 (BMI > 30 kg/m²), and 40 lean (BMI 20-25 kg/m²) subjects (26 men and 14 women) were
190 included. The overweight group had an age range from 37 to 68 years, and the lean from 36 to
191 65 years (Table 1). The data was normally distributed.

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193 **Insulin resistance markers and lipids**

194 Overweight subjects had higher ($P < 0.05$) levels of all insulin resistance markers (insulin,
195 Homeostasis Model Assessment (HOMA), C-peptide, HbA1c) than their lean counterparts.
196 Glucose was elevated ($P = 0.03$) in overweight men relative to the lean ones, but this was not
197 found among the women (Table 1).

198 No significant differences in the plasma concentration of total cholesterol were found
199 between the overweight and lean subjects, but the LDL-cholesterol and triglyceride levels
200 were higher ($P < 0.05$) whereas the HDL-cholesterol level was lower ($P < 0.05$) in the
201 overweight relative to the lean subjects (Table 1).

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203 **Inflammatory markers and adipokines**

204 The overweight subjects had higher ($P < 0.05$) levels of CRP and IL-6 than their lean
205 counterparts. Overweight subjects also had elevated ($P < 0.05$) levels of the leptin and resistin,
206 compared to the lean subjects, while the level of adiponectin was lower ($P < 0.05$). Overweight

207 women had higher levels of CRP than overweight men ($P = 0.05$) (Table 2). and women in
208 both groups had higher ($P < 0.05$) levels of leptin and adiponectin than men (Table 2). No
209 significant difference in plasma levels of IGF-1 was observed.

210

211 **Body composition in overweight and lean subjects**

212 All body composition measures were significantly elevated in overweight compared with lean
213 subjects. Both overweight and lean women had higher TSF ($P < 0.001$), whole body fat (%)
214 ($P < 0.01$) and fat mass ($P < 0.01$) than their male counterparts (Table 3). Males had higher
215 levels for all other measurements than females except for hip circumference, mid upper arm
216 circumference and trunk fat mass.

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218 **Prediction of fat mass**

219 To quantify the relationship between anthropometric estimates of fat mass and body fat
220 measured with BIA we performed multiple linear regression analyses (Table 4). Hip
221 circumference had the highest standardized coefficient and explained most of the variation in
222 whole body- and trunk fat mass (% and kg) in both overweight and lean subjects. Waist-to-hip
223 ratio demonstrated the second highest standardized coefficient for whole body fat mass
224 (% and kg) and trunk fat (%) in overweight subjects. In lean subjects, TSF had the second
225 highest standardized coefficient for all BIA measures of fat mass. In summary, the results
226 showed that measurements of hip circumference were highly associated with whole body- and
227 trunk fat mass expressed in kg and percentage, in both lean and overweight subjects. The
228 results also indicated that an increase in hip circumference with one cm in both overweight
229 and lean subjects corresponded to an increase in the trunk body fat mass with
230 360 g.

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232 **Relationship between insulin resistance markers, lipids and inflammatory markers, and**
233 **body fat**

234 Because measurements of hip circumference were closely related to BIA-derived fat mass in
235 both lean and overweight subjects, tertiles of hip circumference and whole body fat (%) were
236 used to analyse the relation between fat mass and markers of insulin resistance, lipids and
237 inflammatory markers (Tables 5 and 6). We also divided BMI into tertiles and performed the
238 same analysis (Table 7).

239 In overweight subjects, IL-6 was reduced across tertiles of hip circumference
240 (Table 5). Levels of adiponectin and leptin increased, while resistin decreased. There was also
241 an elevation of IGF-1 and CRP concentrations. Levels of HOMA ($P<0.05$) and C-peptide
242 ($P<0.05$) increased and an elevation of triglycerides was seen, while HDL cholesterol
243 remained stable (Table 5). The same trends were found regarding tertiles of BMI in the
244 overweight subjects, except for a significant decrease of resistin ($P<0.05$) and elevated C-
245 peptide ($P<0.01$) levels (Table 7). Across tertiles of whole body fat (%) (Table 6), there were
246 also increasing trends in adiponectin ($P<0.01$) and leptin ($P<0.01$), IGF-1, CRP, HOMA, and
247 C-peptide. Concentrations of IL-6 and resistin ($P<0.05$) increased across tertiles, and
248 triglyceride concentrations decreased.

249 Regarding the relation to tertiles of hip circumference in lean subjects (Table 5), IL-6
250 and adiponectin were reduced, and leptin ($P<0.01$) and resistin values were increased. Levels
251 of IGF-1, CRP, HOMA, C-peptide and triglycerides were increased, while HDL cholesterol
252 was reduced. Similar trends were found for tertiles of BMI in the lean subjects, except for
253 resistin which was decreased across tertiles, and CRP ($P<0.05$), which was significantly
254 increased (Table 7). Like for the tertiles of hip circumference and BMI, leptin ($P<0.01$), IGF-
255 1, CRP, HOMA and C-peptide, increased across tertiles of fat (%) (Table 6). IL-6 values

256 however were stable and resistin ($P < 0.05$) and triglycerides decreased, while HDL cholesterol
257 increased across tertiles of fat (%).

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291 **DISCUSSION**

292 Obesity increases the risk of chronic diseases and the total amount of fat and its distribution
293 are possibly the most important determinants of these disorders (3). Hip circumference was
294 found to be the anthropometric measure that best reflected whole body fat (%) and trunk fat
295 (%) as measured with BIA, in both lean and overweight subjects. Interestingly we found a
296 tendency towards higher concentrations of leptin, CRP, and C-peptide, as well as adiponectin
297 and HDL, with higher fat (%), also in subjects with a BMI within the recommended range.

298 In this study we related frequently used anthropometric measures (BMI, TSF, waist
299 and hip circumference, waist-to-height, waist-to- hip ratio) to fat mass assessed by BIA.
300 Several studies have validated BIA by using DXA (15). In comparison with DXA, BIA tends
301 to overestimate fat mass (% and kg) in lean individuals and underestimate fat mass in obese
302 (17). Despite these limitations BIA is considered an acceptable tool for predicting body fat in
303 healthy populations (11). A recent study also demonstrated that BIA correlated significantly
304 with anthropometric measurements (18). This is in accordance with our study as we found
305 that TSF correlated significantly with BIA measures of fat (% and kg) in lean subjects and
306 waist circumference correlated with BIA-measures of fat in overweight subjects. Hip
307 circumference reflected BIA-measured fat in both groups.

308 In obesity the fat tissue produces adipokines (19) and cytokines, which may result in
309 chronic inflammation (20). It has been shown that systemic inflammation is higher in obese
310 than in normal-weight persons (21). Leptin is preferentially secreted by subcutaneous adipose
311 tissue (22), and the concentration is dependent on adipocyte size (23) as well as energy
312 balance (21). In our study we found a strong association between hip circumference and
313 whole body fat (%), and leptin levels. The same association was also found with BMI.
314 Normally, leptin levels are higher in obese individuals as demonstrated here. Interestingly we
315 observed that leptin levels increased with increasing fat (%) also in the lean group. One could

316 argue that this could be an effect of food intake or macrophage infiltration in adipose tissue
317 due to weight gain, which is known to produce higher leptin levels. However, in both study
318 groups the blood levels were measured during fasting and all subjects reported stable weight
319 for at least two months prior to inclusion. Few studies have shown the same trend with leptin
320 levels in lean people, but a positive association between fat mass accumulation, oxidative
321 stress indices and leptin levels has been observed (7), suggesting that fat mass-induced
322 oxidative stress may cause a dysregulation of adipokines, also in lean subjects.

323 A positive relationship between BMI, waist circumference and CRP has been
324 documented (24). This is in accordance with our study as we found that CRP increased with
325 increasing BMI and interestingly, this positive relationship was significant in lean subjects.
326 We also found an association between hip circumference and whole body fat (%) and CRP,
327 although not significant. These results confirm the findings by Arner et al (25) of an
328 association between inflammation and fat mass in lean individuals. There is also evidence that
329 IL-6, a key determinant of CRP production in hepatocytes, is secreted in proportion to the
330 expansion of fat mass, particularly in the abdominal region (26). We did, however, not detect
331 stronger associations with CRP and trunk fat mass than with other fat measures. Other adipose
332 tissue depots in ectopic sites (liver, heart, skeletal muscle) may contribute to the production of
333 inflammatory mediators in the absence of obesity (27).

334 Chronic inflammation promotes insulin resistance and cardiovascular disease (5). Our
335 results show an increase in HOMA and C-peptide as hip circumference and BMI increased,
336 and an elevation of these markers from the lowest to the highest tertile of whole body fat (%)
337 in both groups. Low level of HDL-cholesterol is an important risk factor for cardiovascular
338 disease (28). One would expect a reduction in HDL-cholesterol as fat mass expands. This was
339 found in our study with increasing hip circumference in lean individuals and with increasing
340 BMI in both groups. In the overweight however, we found stable levels of HDL-cholesterol as

341 hip circumference increased, and elevated levels of HDL-cholesterol from the lowest to
342 highest tertile of whole body fat (%). Elevated HDL-cholesterol levels were followed by an
343 inverse reduction of triglyceride levels. Studies have described a subset of obese individuals,
344 termed metabolically healthy, which appear to be resistant to the development of metabolic
345 disturbances (29). They have high fat mass and high BMI and high HDL, but low
346 triglycerides and visceral fat and normal insulin sensitivity. In our study a subgroup of the
347 overweight people, namely those with BMI > 30 kg/m², but no elevated HOMA, triglyceride-
348 or LDL levels, had the highest levels of whole body fat (%). It should be noted that all the
349 overweight women in our study were in the highest tertile of fat (%). This may also explain
350 our findings regarding adiponectin: In the overweight group we found elevated levels of
351 adiponectin in the highest tertiles of hip circumference, whole body fat (%) and BMI. Earlier
352 studies show a decrease (30) in adiponectin levels as fat mass accumulates and an elevation
353 with weight loss (27).

354 The major strength of our study is that we examined a broad range of anthropometric
355 measures. Our study has some limitations since we used indirect measurements as indicators
356 of total and central fatness. It is therefore difficult to determine exactly the relative
357 contributions of subcutaneous versus visceral fat. The number of subjects was relatively low
358 and the results should be confirmed in a larger population. The age and gender heterogeneity
359 is also a limitation, although the variable was adjusted for.

360 In conclusion, we have showed that measurements of hip circumference to assess total
361 body and trunk fat (%) may represent a valid substitute to BIA measurements in both lean and
362 overweight subjects. Thus this is a highly feasible method outside the hospital setting in order
363 to identify people at risk of increased inflammation and insulin resistance.

364 Our results may also suggest that fat (%) is associated with elevation of risk factors for
365 lifestyle related disorders among lean persons. Although the choice of fat measure may

366 impact on the magnitude of these associations, adherence to a healthy lifestyle is also
367 important for people within the recommended range of BMI. The relationship between
368 markers of inflammation, insulin resistance and lipids in lean as well as overweight subjects
369 should be studied further in order to understand the role of fat mass in healthy subjects with
370 different BMI. Such knowledge may be of considerable interest for early identification of
371 subjects at risk of type 2 diabetes and cardiovascular disease.

372

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374 **Statement of authorship**

375 NWR, VHTH, SMU and AB were responsible for the original ideas and methodology of the
376 Study, which was conducted by NWR, VHTH and IN. The blood samples were analysed by
377 VHTH and IN. NWR, KBH and AB performed the statistical analyses. Financial support was
378 obtained by MJH and SMU. NWR, KBH, ID, POI, MJH, SMU and AB were responsible for
379 the data interpretation, and discussions regarding drafting of the manuscript. All co-authors
380 have made substantial contributions in the writing process and approved the final manuscript.

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382 **Conflict of interest**

383 The authors declare no conflict of interest.

384

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